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Correlation between oxidative stress and Hydroxyproline content in Liver Fibrosis

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ADTICLE INFO	

ARTICLE INFO	
Article history:	Background & Objectives: Hepatic fibrosis is the extreme
Received:	extracellular matrix buildup of collagen due to the chronic liver
Accepted: 16/3/2021	disorders. Progressive liver fibrosis results in cirrhosis and
Available online :	hepatic failure. Nilotinib is considered a 2^{nd} generation tyrosine
	kinase inhibitor which exhibited antifibrotic efficacy Stem
Keywords:	cells release cytokines and growth factors which exhibited
Hepatic fibrosis Nilotinib	paracrine_mediated antifibratic and anti-inflammatory effects
	in vitro and in vivo. Honco, stam calls conditioned medium
	In vitro and in vivo. Hence, stem cens conditioned medium
	(SC-CM) may have an antifibrotic role. This study is designed
	to explain the association between the oxidative stress markers
	and the hydroxyproline content in liver fibrosis. Methods:
	Sixty male wistar rats were distributed into six major groups;
	control group, CCl ₄ group, Free conditioned media group,
	(CCl ₄ & Nilotinib) group, (CCl ₄ & stem cells exosomes) group,
	and (CCl ₄ & Nilotinib & stem cells exosomes) group. They had
	an intraperitoneal injection with CCl ₄ for nine weeks and had
	treatments of Nilotinib, stem cell exosomes (extracted from
	Liver mesenchymal stem cells), and a combined treatment of
	Nilotinib and stem cell exosomes from the 5 th week to the 9 th
	week of CCl ₄ intoxication. Oxidative stress parameters (e.g.
	MDA, SOD, GSH, CAT, NO, and NOS) and hydroxyproline
	content were estimated for each group Results: results showed
	a significant positive correlation between Hydroxyproline and
	MDA NO and NOS Also there is a significant negative
	appropriate hot was ho
	Conclusion: There is a significant positive correlation between
	Unclusion. There is a significant positive correlation between
	Hydroxyproine and MDA, NO and NOS. And there is a
	significant negative correlation between Hydroxyproline and
	GSH, SOD, CAT.

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INTRODUCTION

Fibrosis is a prevalent pathological progression for the most of liver disorders

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that results in hepatic cirrhosis and hepatocellular carcinoma (HCC). It results from nearly all chronic liver disorders mainly rising from metabolic. viral. autoimmune disorders [1]. Fibrosis is a consequence of chronic wound healing and is described as the advanced replacement of efficient liver tissue with extracellular matrix which is rich with collagen I and III; it disturbs the normal construction and functions of the liver particularly in the late stage of liver cirrhosis. Liver fibrosis is also believed a precancerous condition which supplies microenvironments where 1ry tumors may progress [2].

Oxygen free radicals act as а significant part in tissue destruction in many liver diseases. Free radicals like the hydroxyl radicals and superoxide, can give rise to a destruction in cellular constituents by peroxidation of lipoproteins and nucleic acids [3]. Peroxidation is initiated by free radicals which leads to oxidative destruction of polyunsaturated fatty acids of cell membranes and also leads to making aldehyde metabolites such as malondialdehyde (MDA) [4].

Another study reported that (Nitric oxide) NO plays a role in liver fibrosis. NO activates guanylyl cyclase (GC) by connecting to its metal center. This forms cyclic guanosine monophosphate (cGMP) that connects to cGMP-dependent protein kinases (PKGs) and enable several biological processes like platelet aggregation, vasodilation and neurotransmission. Inducible nitric oxide synthase (iNOS)-derived NO is implicated in the growth and maintenance of many liver diseases [5]. Also there is a study reported that the actions of superoxide dismutase (SOD), Catalase (CAT) were found to be lower in cirrhotic liver in comparison with healthy liver. Those changes were related to a reduction of reduced glutathione (GSH) levels in cirrhotic liver tissue in comparison with healthy liver tissue [6]. Furthermore, it was reported that the apoptosis of hepatocytes causes the induction of the inflammatory cells to the injured liver and discharge of profibrogenic cytokines such as TGF- β 1. TGF- β 1 is crucial to activating the fibrogenic myofibroblasts, which release extracellular matrix proteins like collagen Type I and hydroxyproline [**7**].

The present study is designed to explain the correlation between oxidative stress and hydroxyproline in the advancement of liver fibrosis.

Material and Methods: Experimental Animals:

Fifty male Wistar rats (180-200 gm) purchased from the Egyptian were Organization for Biological Products and Vaccines (VACSERA, Giza, Egypt). The rats were housed at $21\pm2^{\circ}$ C, $50\pm5\%$ relative humidity. Rats were preserved in cages at a steady environment and nutritional condition during the experiment. All the procedures associated with animal care, handling, treatment and scarification are firmly followed to the Guide for the Care and Use of Laboratory Animals published by the US National Institute of Health (Publication No.85-23, revised 1996).

Study Design:

Rats were distributed into six main groups (10 rats in each cage) as follows. Group 1: rats were injected with 1 ml/kg of corn oil for nine weeks. Group 2: rats had intraperitoneal injection of 1 ml/kg of CCl4 solution for nine weeks [8]. Group 3: rats were injected with 1 ml/kg of CCl4 solution intrperitoneally+ exosome free media (1ml daily) from the 5th week to the 9th week of CCl4 intoxication. Group 4: rats had an intraperitoneal injection with 1 ml/kg of CCl₄ solution+ Nilotinib (20 mg/kg daily) (formerly AMN107; Tasigna[®]) from the 5th week to the 9th week of CCl₄ intoxication [9]. Group 5: rats were injected with 1 ml/kg of CCl₄ solution intraperitoneally + 1ml of stem cell exosomes/rat daily (From liver mesenchymal cells (10000)stem cells/cm²) from the 5th week to the 9th week of CCl₄ intoxication [10]. Group 6: rats were injected with 1 ml/kg of CCl_4 solution intraperitoneally+ stem cell exosomes (1 ml daily) + Nilotinib (20 mg/kg daily) from the 5th week to the 9th week of CCl_4 intoxication.

Biochemical assays:

Rats from the six groups were sacrificed and a general anesthesia called before thiopental were used the sacrification. Α laparotomy and hepatectomy were done. The liver samples were kept in liquid nitrogen. GSH, NO, MDA. NOS. CAT. SOD and hydroxyproline were assessed by using BioVision reagent kits, USA. The tests were done according to the kit instructions [11] [12][13][14][15][16].

Statistical analysis:

The Statistical Package for the Social Sciences (IBM SPSS, version 20) was applied for this study to analyze the statistics of the results. One-way ANOVA was made and the numbers were obtained as mean \pm SD. A simple linear correlation was analyzed by Pearson's method for measuring the degree of dependency between variables (IBM SPSS).

Results:

Oxidative stress markers among the studied groups:

The administration of CCl₄ gave rise to a significant elevation of NO, MDA and NOS in comparison with Group 1. Nilotinib and stem cells exosomes treatments significantly reduced the NOS and NO in comparison with Group 2. Stem cells exosomes treatment faintly reduced the elevated levels of MDA in comparison with Group 2, but the Nilotinib treatment gave rise to a significant reduction on MDA in comparison with Group 2. The combined treatment of Nilotinib and stem exosomes cells displayed a further significant decrease in NO, MDA and NOS than the other treatments. This study also showed no significance in NO, MDA and NOS between Group 2 and Group 3. The CCl4 administration gave rise to a significant reduction in hepatic CAT, GSH and SOD in comparison with Group 1. Nilotinib and stem cells exosomes treatments significantly raised the SOD and GSH in comparison with Group 1. Stem cells exosomes treatment faintly raised the levels of CAT in comparison with Group 1, but the Nilotinib treatment gave rise to a significant elevation on CAT comparison with Group in 2. The combined treatment of Nilotinib and stem cells exosomes revealed а further significant increase in CAT. GSH and SOD than the other treatments. There was no significance in CAT, GSH and SOD between Group 1 and Group 2 (Table 1).

Hydroxyprotine content among the studied groups

The administration of CCl₄ gave rise significant elevation of to а Hydroxyproline in comparison with Group 1. Nilotinib treatment significantly reduced the Hydroxyproline content in comparison with Group 2, but the stem cells exosomes treatment faintly reduced the Hydroxyproline content in comparison with Group 2. The combined treatment of Nilotinib and stem cells exosomes displayed a further significant reduction in the Hydroxyproline content than the other treatments. This study also showed no significance in the levels of hydroxyproline content between Group 2 and Group 3 (Table 1).

Correlation between Hydroxyproline content and all markers:

There is a significant positive correlation between Hydroxyproline and MDA, NO and NOS. And there is a significant negative correlation between Hydroxyproline and GSH, SOD, CAT (Figure 1) (Table 2).

Discussion:

This present study revealed that combined treatment of Nilotinib and stem cells exosomes had a significantly greater antifibrotic outcome than each treatment alone. There were significant declines of hydroxylproline content and oxidative stress in the combined treatment group (Group 6) in comparison with each treatment alone.

The antifibrotic effect of combined treatment of nilotinib and stem cells exosomes may be owing to a synergic effect of the 2 types of treatment: Stem cells exosomes have anti-inflammatory impact and decrease iniuries to hepatocytes that inhibit the hepatic stellate cells (HSCs) activation which results in HSCs apoptosis and fibrinolvsis stimulation [17]. Otherwise, MSCs alter the macrophages' polarity to an antiinflammatory phenotype. elevate the making of matrix metalloproteinases for reducing the ECM, and increasing the hepatocyte phagocytosis of remains (throughout this process, macrophages elevate the pro-regenerative factors) [18]. Also. Nilotinib has antifibrotic effect through 3 main ways tangled in fibrogenesis: deactivation of c-Abl tyrosine kinase which prompt TGF- β leading to fibrogenesis, suppression of discoidin domain receptors and collagen receptors [9], thus MSCs exosomes cause fibrinolysis and Nilotinib causes fibrinogenesis.

This study showed a significant between positive correlation Hydroxyproline and MDA, NO and NOS. Also, there is a significant negative correlation between Hydroxyproline and GSH, SOD and CAT. Gabr et. al, reported that hydroxyproline is about 1/3 of the amino acids in the collagen. Fibrosis activation depend mostly on the collagen fibers deposition. It is associated with over deposition of ECM, because of inequality between synthesis and degradation of in hepatic tissues [19]. NOS ECM produces NO from L-arginine in a reaction which converts arginine and oxygen into citrulline and NO, which in turn NO is capable adjusting the matrix of metalloproteinases' activity and the formation of constituents of ECM like the hydroxyproline [20] [21][22][23] and that's why there is a positive correlation between hydroxyproline and NO (or NOS).

Also, there is a studies reported that GSH plays an important role in detoxification within cells, mostly cells of the intestines, kidney, lungs and liver. GSH has a significant relationship with lipid peroxidation due to the ability of these molecules to combine with free radicals that start lipid peroxidation and reduced hydrogen peroxide formed inside cells. Lipid peroxidation is initiated by free radicals which leads to oxidative damage of polyunsaturated fatty acids which are constituent of cell membranes and also results in the production of reactive aldehyde compounds such as MDA [4][24]. And that's the reason for positive correlation between hydroxyproline and MDA and a negative correlation between hydroxyproline and GSH.

Also Arya et. al., reported that SOD is one of the key enzymes which defend the body against oxidative stress. This enzyme catalyzes conversion of 2 O_2^- to H_2O_2 and O_2 . H_2O_2 can be changed into O_2 and H₂O by CAT [25]. Shin et al., reported that CAT participates in the protection of mitochondria against exogenous or endogenous H_2O_2 which causes lipid peroxidation for the hepatocytes [26]. And that's why there is a negative correlation between hydroxylproline and CAT (or SOD).

Conclusion:

Combined treatment of Nioltinib and stem cells exosomes was more antifibrotic than each treatment alone. Also, Oxidative stress markers were associated with the hydroxyproline content in liver fibrosis because there is a significant positive correlation between Hydroxyproline and MDA, NO and NOS. Also, there is a significant negative correlation between Hydroxyproline and GSH, SOD and CAT.

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Groups	Hydroxyproline content (measure of fibrosis)(ug/mg	Malonaldehyde (mmol / g tissue)	Nitric oxide (micro mol /g tissue)	Glutathione reduced (micromol/g protein)	Super oxide dismutase (U/mg protein)	Nitric oxide synthase (P MO L/MIN/mg protein)	Catalase (mol / min / gm)
	liver						
Control	31.21±3.76	77.93±5.88	150.84±19.25	23.47±3.56	21.09±2.86	18.03±2.61	1.05 ± 0.11
CCl4	61.60±21.56	111.17±18.21	205.51±25.30	14.78±3.02	14.11±3.96	28.36±5.01	0.66±0.24
Free media	65.30±17.85	110.71±16.80	205.76±26.19	14.51±3.45	14.37±3.82	29.27±3.26	0.64±0.22
CCl4+Nilotinib	44.04±10.18	83.97±14.78	176.39±22.37	23.85±5.08	19.70±4.42	20.14±5.66	0.99±0.21
CCl4+stem cells exosomes	55.77±15.78	96.480±28.41	173.25±26.41	21.32±3.37	18.33±3.25	20.04±2.85	0.80±0.26
CCl4+stem		80.10±17.83	148.02±14.02	23.28±5.24	22.80±3.82	18.98±3.27	1.02±0.21
cells exosomes + Nilotinib	34.88±5.19						
p-value	<0.001*	<0.001*	<0.001*	<0.001*	< 0.001*	< 0.001*	< 0.001*

Table 1: Oxidative stress markers and hydroxyprotine content among the studied groups:

Table 2: Correlation between the oxidative stress markers and hydroxyproline content:

		Hydroxyproline content (ug/mg liver)
МДА	r	0.272
MDA	р	0.036*
CSH	r	-0.528
6511	р	<0.001*
NO	r	0.346
NU	р	0.007*
NOS	r	0.385
NUS	р	0.002*
SOD	r	-0.599
300	р	<0.001*
САТ	r	-0.463
UAI	р	<0.001*



Figure (1): Correlation between the oxidative stress markers and hydroxyproline content